

BUSTER reference card	
set up	
Set up for csh or tcsh	source /some/where/buster/installed/setup.csh
Set up for sh, bash, ksh, or dash	. /some/where/buster/installed/setup.sh
Check all 3rd party tools used by BUSTER work	checkdeps
Download PDB entry (code 1abc) and convert SF to mtz	fetch_PDB 1abc
BUSTER refinement most useful options	
Brief help message	refine -h
Default refinement: long 5 big cycles * 100 small cycles	refine -p protein.pdb -m reflects.mtz \ -d res.dir > results.log
No refinement, just maps	refine -p pdb -m mtz -d dir -M MapOnly > log
Quick refine (after coot rebuild)	refine -p pdb -m mtz -d dir -M ShortRunVoid > log
Medium length refinement	refine -p pdb -m mtz -d dir -nbig 2 > log
Use NCS restraints (LSSR)	refine -p pdb -m mtz -d dir -autoncs > log
TLS basic mode	refine -p pdb -m mtz -d dir -M TLSbasic > log
Ligand dictionary (see below)	refine -p pdb -m mtz -d dir -l grade-XXX.cif > log
Use QM method for ligand LIG	refine -p pdb -m mtz -d dir -l grade-LIG.cif -qm LIG \ > log
Rigid body for first big cycle	refine -p pdb -m mtz -d dir -RB > log
Water updating	refine -p pdb -m mtz -d dir -WAT > log
Target restraints (LSSR) to a reference structure	refine -p pdb -m mtz -d dir -target high_res.pdb > log
Occupancy refinement	pdb2occ -p pdb -o pdb-occ.Gelly refine -p pdb -m mtz -d dir -Gelly pdb-occ.Gelly > log
Find ligand binding sites for rhoFit	refine -p pdb -m mtz -d dir -L > log
Add hydrogen atoms to protein and its ligand, then refine. Use at better than 2.0Å resolution	hydrogenate -p prot.pdb -l grade-XXX.cif -o protH.pdb refine -p protH.pdb -m mtz -d dir -l grade-XXX.cif > log
An example: initial very long refinement of molecular replacement solution	refine -p MR.pdb -m refl.mtz -d res.dir \ -RB -target MR.pdb -nbig 10 > results.log
Looking at BUSTER refinement results	
Produce report on refine run	buster-report -d dir
view buster-report output	firefox dir-report/index.html
Start coot to see structure with maps plus “unhappy atoms” list	visualise-geometry-coot res.dir

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BUSTER reference card	
cif restraint dictionary preparation	
Brief help message	grade -h and grade_PDB_ligand -h
SMILES residue name LIG note: use of ''	grade -resname LIG 'C1CN(CCN1CCO)CCS(=O)(=O)O'
From mol2 file (with hydrogen atoms)	grade -in ligand.mol2 -resname LIG
Charged ligand	grade -in acid.mol2 -resname LIG -charge -1
Ligand exists in PDB eg 3AS	grade_PDB_ligand 3AS
Examine (and edit) cif dictionary	EditREFMAC grade-LIG.cif grade-CUF.pdb LIG
Ligand fitting	
Brief help message	rhofit -h
Fit ligand XXX	rhofit -p protein.pdb -m refine.mtz -l grade-XXX.cif \ -d rhofit.dir
Fit ligand in 2 sites	rhofit -p prot.pdb -m mtz -l cif -d dir -xclusters 2
Allow chiral centres to invert during fit	rhofit -p prot.pdb -m mtz -l cif -d dir -nochirals
Look at rhofit results using coot	cd rhofit.dir visualise-rhofit-coot
refine from rhofit best fits in each site (use with caution!)	refine -p rhofit.dir/merged.pdb \ -l rhofit.dir/best.cif -m mtz -autoncs > log
Other tools	
Calculate an anomalous Fourier map	diff_fourier -m truncate.mtz -p refine.mtz \ -P PH2FOFCWT FOM
Calculate a Fo-Fo difference Fourier map (and compare positive peaks to PDB file)	diff_fourier -m apo.mtz -m2 inhibitor.mtz \ -p apo_refine.mtz -P PH2FOFCWT FOM \ -o Fo-Fo -noANO \ -pdb apo_refine.pdb
Calculate an early-late difference Fourier map (and compare within 3Å to PDB file)	diff_fourier -m late.mtz -m2 early.mtz \ -p refine.mtz -P PH2FOFCWT FOM \ -o early-late -noANO \ -pdb refine.pdb -compare_cut 3.0

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